# ANTITUMOR AGENTS, 81.<sup>1</sup> JUSTICIDIN-A AND DIPHYLLIN, TWO CYTOTOXIC PRINCIPLES FROM *JUSTICIA PROCUMBENS*

NARIHIKO FUKAMIYA and KUO-HSIUNG LEE\*

#### Natural Products Laboratory, Division of Medicinal Chemistry and Natural Products, School of Pharmacy, University of North Carolina, Chapel Hill, North Carolina 27514

The whole plant of Justicia procumbens L. (Acanthaceae), known as "Chu Wei Hung" or "Jué Chuáng" in Chinese folklore, is used as an herbal remedy for the treatment of fever, pain due to pharyngo-laryngeal swelling (1-3), and cancer (4). Prior phytochemical studies on this plant have afforded justicidin-C (4) and justicidin-D (3) (5). As a result of our continuing searches among Chinese medicinal plants for novel, naturally occurring potential antitumor agents (6), the methanolic extract of the whole plant of J. procumbens was found to show significant inhibitory activity in vivo against P-388 lymphocytic leukemia growth in  $BDF_1$  male mice (T/ C=150% at 50 mg/kg/day, IP) as well as in vitro cytotoxicity in the 9-KB (human nasopharyngeal carcinoma) cell culture assay (7). Bioassay-directed fractionation of the foregoing MeOH extract led to the isolation of five 2,3-naphtalide lignans, which were identified as justicidin-A (1), diphyllin (2), neojusticin-A (i.e., justicidin-D) (3), neojusticin-B (i.e., justicidin-C) (4), and justicidin-E (5) by spectral analysis and direct comparison with authenic samples of the first four compounds (8,9) as well as data for the last compound reported in the literature (10). Compounds 1-4 were previously isolated from J. procumbens var. leucantha (8,9). Compound 1 was also reported as a fish-killing component of Justicia hayatai var. decumbens (11).

A comparison of the cytotoxicity (KB) clearly indicated that the  $\gamma$ -lactone ring carbonyl  $\alpha$  to C-3 instead of C-2 is re-

quired for potent cytotoxicity. Thus, 1, and 2, were active  $[ED_{50} (KB) < 1.0 \ \mu g/$ ml each],<sup>2</sup> while **3-5** were inactive  $[ED_{50} (KB)=9.0 \ \mu g/ml$  for **3** and >10.5  $\mu g/ml$  for **4** and **5** each].

# **EXPERIMENTAL**

GENERAL EXPERIMENTAL PROCEDURES.— Melting points were determined on a Thomas-Hoover melting point apparatus and are uncorrected. Ir spectra were recorded on a Perkin-Elmer 257 grating spectrophotometer. <sup>1</sup>H-nmr spectra were recorded on a Bruker WM-250 Fourier Transform spectrometer and are given in parts per million ( $\delta$ ) downfield from an internal TMS standard. The abbreviations s and d refer to singlet and doublet, respectively. Mass spectra were determined on an A.E.I. MS-902 instrument at 70 eV using a direct inlet system. Silica gel for column chromatography refers to Merck silica gel 60 (70-230 mesh). Silica gel for preparative tlc refers to Analtech silica gel G (1000 microns). Compounds were visualized by uv light or spraying with 1% Ce(SO<sub>4</sub>)<sub>2</sub>-10% H<sub>2</sub>SO<sub>4</sub> solution followed by heating.

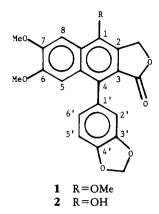
PLANT MATERIAL.—The *J. procumbens* used was from a collection made in the spring of 1979 in Jui-Li, Chia-Yi-Shen, Taiwan, by Professor Huan-Chang Huang. A voucher specimen (HCH-73) is available for inspection at the herbarium of the School of Pharmacy, Kaohsiung Medical College, Kaohsiung, Taiwan.

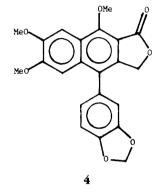
PRELIMINARY EXTRACTION.—The ground, air-dried whole plant (2.27 kg) was exhaustively extracted with hexane and then MeOH. The MeOH extract (132 g) was partitioned between CHCl<sub>3</sub> and H<sub>2</sub>O. The aqueous layer was extracted with BuOH. The hexane, *n*-BuOH, and the aqueous layer were not examined further as they were not active in the in vitro KB assay.

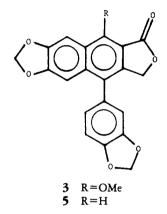
Isolation of Justicidin-A (1), Diphyllin (2), NeoJusticin-A (3), NeoJusticin-B (4),

<sup>&</sup>lt;sup>1</sup>For Part 80, see G.K. Rice, T. Yokoi, T. Hayashi, H. Suzuki, A.T. McPhail, and K.H. Lee, J. Chem. Soc. Chem., Commun., (submitted).

<sup>&</sup>lt;sup>2</sup>The cytotoxic lignans isolated from *J. procumbens* are **1** and **2**, instead of **3** and **2** as reported by Lee (6), p. 361. The structure of diphyllin reported as **30** in the foregoing References 6 was erroneous and has to be revised to **2**.







AND JUSTICIDINE-E (5).—The KB active CHCl<sub>3</sub> extract (36.1 g) was chromatographed on silica gel (2.3 kg) and eluted with CHCl<sub>3</sub> (15 liters), Me<sub>2</sub>CO (8 liters), and MeOH (8 liters). All fractions were monitored by the foregoing KB cytotoxicity assay. The active CHCl<sub>3</sub> fractions (4.2 g) were combined and rechromatographed on silica gel (1.5 kg). Elution of the column with C<sub>6</sub>H<sub>6</sub> and C<sub>6</sub>H<sub>6</sub> with increasing amount of EtOAc gave six active fractions. Purification of these fractions with preparative tlc [silica gel, CHCl<sub>3</sub>-Me<sub>2</sub>CO (5:1)] yielded five lignans (1-5)<sup>3</sup>, in which 1 and 2 were cytotoxic while 3-5 were inactive as mentioned above.

Compound 1 (8.1 mg, 0.00036% yield): mp 261-263°; Okigawa *et al.* (8) reported mp 261-263°; ir (CHCl<sub>3</sub>) 1758 and 1615 cm<sup>-1</sup>; <sup>1</sup>H nmr (CDCl<sub>3</sub>)  $\delta$  3.80 (3H, s) 4.08 (3H, s), 4.12 (3H, s) (3 x OMe), 5.55 (2H, s, CH<sub>2</sub>OCO), 6.04 (1H, d, J=1.0 Hz), 6.06 (1H, d, J=1.0 Hz) (OCH<sub>2</sub>O), 6.77 (1H, dd, J=8.0 and 2.0 Hz, H-6'), 6.81 (1H, overlapped d, J=2.0 Hz, H-2'), 6.95 (1H, d, J=8.0 Hz, H-5'), 7.09 (1H, s, H-8) and 7.55 (1H, s, H-5). Compound 1 was identical by tlc, mmp, ir, and nmr spectra with an authenic sample of justicidin-A.

<sup>3</sup>Compounds **1-5** showed Rf values 0.53, 0.25, 0.58, 0.53, and 0.47, respectively, on silica gel in  $C_6H_6$ -EtOAc (6:1).

Compound 2 (3.2 mg, 0.00014% yield): mp 284-287°; Okigawa *et al.* (8) reported mp 284-287°; ir (KBr) 3300 and 1730 cm<sup>-1</sup>; <sup>1</sup>H nmr Me<sub>2</sub>CO- $d_6$ )  $\delta$  3.71 (3H, s), 3.97 (3H, s) (2 x OMe), 6.05 (2H, s-like, OCH<sub>2</sub>O) and 7.67 (1H, s, H-5). This compound was identical with an authenic sample of diphyllin by direct comparision (tlc, mmp, ir, and nmr spectra).

Compound **3** (12.9 mg, 0.00057% yield): mp 273-275°; Okigawa *et al.* (8) reported mp 273-275°; ir (CHCl<sub>3</sub>) 1755 and 1600 cm<sup>-1</sup>; <sup>1</sup>H nmr (CDCl<sub>3</sub>)  $\delta$  4.35 (3H, s, OMe), 5.15 (2H, d, J=3.2 Hz, CH<sub>2</sub>OCO), 6.09 (4H, m, two OCH<sub>2</sub>O, 6.72 (1H, dd, J=8.0 and 2.0 Hz, H-6'), 6.78 (1H, overlapped d, J=2.0 Hz, H-2'), 6.95 (1H, d, J=8.0 Hz, H-5'), 7.00 (1H, s, H-8) and 7.69 (1H, s, H-5). The identity of **3** with an authenic sample of neojusticin-A was established by direct tlc, mmp, ir, and nmr spectral comparison.

Compound 4 (10.2 mg, 0.00045% yield): mp 262-265°; Okigawa *et al.* (8) reported mp 262-265°; ir (CHCl<sub>3</sub>) 1760 and 1610 cm<sup>-1</sup>; <sup>1</sup>H nmr (CDCl<sub>3</sub>)  $\delta$  3.84 (3H, s, C<sub>7</sub>-OMe), 4.07 (3H, s, C<sub>6</sub>-OMe), 4.32 (3H, s, C<sub>1</sub>-OMe), 5.15 (2H, s, CH<sub>2</sub>OCO), 6.08 (1H, d, J=1.0 Hz), 6.09 (1H, d, J=1.0 Hz) (OCH<sub>2</sub>O), 6.80 (1H, dd, J=8.0 and 2.0 Hz, H-6'), 6.83 (1H, overlapped d, J=2.0 Hz, H-2'), 6.90 (1H, d, J=8.0 Hz, H-5'), 6.98 (1H, s, H-8) and 7.69 (1H, s, H-5). This compound was identical with an authenic sample of neojusticin-B by direct comparison (tlc, mmp, ir, and nmr).

Compound 5 (4.5 mg, 0.0002% yield): mp 271-272°; Wada et al. (10) reported mp 265-269°; ir (CHCl<sub>3</sub>), 1758 ( $\gamma$ -lactone) and 1598 (aromatic ring) cm<sup>-1</sup>; <sup>1</sup>H nmr (CDCl<sub>3</sub>)<sup>4</sup>  $\delta$  5.19 (2H, d, J=3.2 Hz, CH<sub>2</sub>OCO), 6.08 (4H, m, two OCH<sub>2</sub>O), 6.77 (1H, dd, J=8.0 and 2.0 Hz, H-6'), 6.81 (1H, overlapped d, J=2.0 Hz, H-2'), 6.98 (1H, d, J=8.0 Hz, H-5'), 7.10 (1H, s, H-8), 7.30 (1H, s, H-5) and 8.24 (1H, s, H-1); ms m/z 348.0639 (M<sup>+</sup>, 100%; calcd for C<sub>20</sub>H<sub>12</sub>O<sub>6</sub>: m/z 348.0632). The comparable melting point of 5 with justicidin-E, coupled with the foregoing spectral data which are consistent with structure 5, led to the assignment of 5 as justicidin-E (12).

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<sup>&</sup>lt;sup>4</sup>The different chemical shifts and splitting pattern observed between **5** and neojusticidin-E reported in Reference 10 might be due to the use of different solvents and instruments, in which the former was run in  $CDCl_3$  at 250 MHz whereas the latter was measured in DMSO at 100 MHz instrument.